

A Large Cohort Study of Genotype and Phenotype Correlations of Beta- Thalassemia in Iranian Population

Fereshteh Maryami¹, Azita Azarkeivan², Mohammad Sadegh Fallah³, Sirous Zeinali^{1,3}

¹Biotechnology Research Center, Department of Molecular Medicine, Pasteur Institute of Iran, Tehran, Iran

²Pediatric Hematology Oncology, Transfusion Research center, High Institute for Research and Education in Transfusion Medicine, Department of Thalassemia Clinic, Tehran, Iran

³Kawsar Human Genetics Research Center, Tehran, Iran

Corresponding Author: Sirous Zeinali, PhD. Iranian Molecular Medicine Network, Biotechnology Research Center, Pasteur Institute of Iran, Pasteur St, Tehran, Iran

Tel and Fax: +98 218 893 9142

Email: sirouszeinali@yahoo.com

Received: 16, Dec, 2014

Accepted: 27, Jan, 2015

ABSTRACT

Background: Thalassemia syndromes are the most prevalent single gene disorders in Iran. This study aimed to evaluate the effect of different types of beta-globin gene mutations, co-inheritance of alpha-globin gene mutations and/or Xmn1 SNP on disease phenotype in a large cohort of Iranian patients.

Subjects and Methods: In total, 433 patients were clinically classified into β -thalassemia major (TM) or intermedia (TI). Multiplex PCR, ARMS-PCR, RFLP-PCR and DNA sequencing were performed to identify both α - and β -globin gene mutations and Xmn1 polymorphism as well. All data were compared and analyzed by SPSS software in TM and TI groups, consequently.

Results: A total of 39 different β -globin mutations were identified. Among them, the most common were IVS IInt1 (40.33%) followed by IVS Int5 (9.56%), C30 (7.22%) and Fr8-9(7%). All patients were subjected to evaluate common α -globin gene deletions. The patients inherited concomitant mutations of α - and β -globin, showed no clinical modifications compared with those who had only β -globin mutation. The TI patients showed a significant increase in frequency of both heterozygous and homozygous form of the Xmn1 polymorphism. It was also found that β^0/β^0 genotype patients, inherited the Xmn1 polymorphism required lesser blood transfusion.

Conclusion: No significant differences were observed, on the severity of disease, between patient's inherited defective α - and β -globin genes and ones with just β -globin gene mutation. Taking the results of this research into account, Xmn1 polymorphism can be considered as an important genetic factor modulating the severity of disease.

Keywords: α -thalassemia, β -thalassemia major, β -thalassemia intermedia, Xmn1 polymorphism, Iran

INTRODUCTION

Although, thalassemia is the most common monogenic disorders in Iran, but it is a very heterogeneous disease at the molecular and clinical levels.¹ These variations depend on the extent of imbalances created between α - and non- α -globin chains synthesis.²⁻³ The incidence of β -thalassemia (β -thal) in Iran has been significantly decreased since 1997 due to the implementation of the National Program for the Prevention of

Thalassemia.⁴⁻⁶ An average fall of about 81% has been reported for 2007-2009.⁷ The last available data about living patients, dated on 2007, revealed 13,879 thalassemia patients, with the mean age of 15 years old, all over the country.⁶ The carrier frequencies of β - and α -thalassemia (α -thal) are estimated to be 4-8%⁸ and 30%,⁹ respectively. Diagnosis of β -thal trait is suggested based on complete blood count (CBC) indices and hemoglobin (Hb) A2 level, although multiple factors

like iron deficiency or α -thal trait should be taken into account.¹⁰ Moreover, It is claimed that co-inheritance of α - with β -hemoglobinopathies remarkably influences the clinical and hematological features of the disease. The later, might be related to the amount of α -globin chain deficiency which is associated with the type of the gene mutation.¹¹

A wide range of α - and β -thal mutations has been detected among Iranian population. The most common α -globin gene mutation is $-\alpha^{3.7}$ kb^{1,12} followed by $-\alpha^{4.2}$ kb, $-\alpha^{20.5}$ kb and --Med deletions.¹³ Alpha-globin gene deletions account for more than 60% of α -globin mutations.¹⁴⁻¹⁶

The IVS IIInt1 is reported to be the most common beta-globin gene mutation in Iran, followed by other point mutations depending on the population ethnicity.^{6,17-18} The frequency of α - and β - globin gene mutations vary throughout Iran notably from north to south.¹⁹⁻²¹

In previous studies, Xmn1-158 γ (C \rightarrow T) (Xmn1 polymorphism) variant has been found to have an increasing effect on the HbF level, ameliorating the severity of the disease.²²⁻²³

This study aimed to evaluate factors affecting genotype and phenotype correlations in thalassemia major or intermedia such as co-inheritance of α -globin gene mutations and/or Xmn1 polymorphism in a large cohort study of Iranian β -thalassemic population.

SUBJECTS AND METHODS

Subjects

A total of 433 patients suffering from either β -thal major or intermedia were admitted into this study. They were 200 males and 233 females; 32 children (≤ 12 Y) and 401 adults (> 12 Y). Patients were referred to Kawsar Human Genetics Research Center by qualified hematologists from Thalassemia Clinics (Ethical committee No: 86022/14). According to clinical status, patients were classified into two groups: thalassemia major (TM) (n=89) and thalassemia intermedia (TI) (n=255).

The remaining patients (n=89) did not have any documented diagnosis. After obtaining informed consent, about 10 ml blood samples were collected in EDTA. DNA extracted via the standard salting out

method,²⁹ were quantified by nano drop spectrophotometer.

DNA genotyping ARMS-PCR method was employed to detect common β -globin genes mutations.³

DNA sequencing was done by Kawsar Biotech Co, Tehran, Iran (KBC), using BigDye Terminator Kit (Thermo Fisher Scientific Inc. Foster City, CA, USA, TF) and the samples were run on ABI 3130XL Genetic Analyzer (TF).

Common α -globin gene deletions ($-\alpha^{3.7}$, $-\alpha^{4.2}$, $-\alpha^{20.5}$ and --MED) were detected using a multiplex gap PCR method.²⁵ Alpha-triplication was detected either by using PCR.²⁶ Xmn1 polymorphism was investigated using RFLP-PCR method.²⁷

Statistical analysis

All statistical analysis was performed with SPSS (version 16) (SPSS Inc., Chicago, IL, USA). Nonparametric Mann-whitney U test was used to compare TM and TI groups. Associations of β -thal with α -thal deletions and/or Xmn1 polymorphism were analyzed by nonparametric Chi-square and Kruskal-wallis (KW) test.

RESULTS

A variety of factors were considered in our study design. Association of each factor with two groups of patients was taken into consideration. Table 1 provides a brief summary of measurements.

DNA samples were analyzed for α - and β -globin gene mutations by various molecular methods. A total of 39 different β -globin mutations were determined. IVS IIInt1 was the most common detected mutation (40%) followed by IVS Int5 (10%), C30 (7%), and Fr8-9 (7%). The allele frequencies for different mutations are shown in Table 2.

Considering β -globin genotypes (i.e. β^{++} , β^{+} and β^{0}), patients were classified into different groups in accordance with criteria provided by Weatherall and Clegg.³ The data is presented in Table 3.

DISCUSSION

According to the genotyping studies, the IVS IIInt1 (G \rightarrow A) was found as the most common mutation. All identified β -globin gene mutations (Table 2) are in a good agreement with most of the previous studies.

Table 1: Clinical and molecular information of the β -thal patients

	No (%)	Transfusion dependency (%)	Mean age at first BT (months)	Transfusion interval (days)	Splenectomy status (%)	Hydroxyurea treatment (%)	BT interruption after treatment (%)	Xmn1 polymorphism			Alpha genotype		
								+/+ (%)	+/- (%)	-/- (%)	Mild α -thal (%)	Severe α -thal (%)	H disease (%)
TM	89 (25.7)	85 (95.5)	24 \pm 21	27 \pm 24	39 (89.6)	4 (4.5)	0	12 (13.4)	37 (41.5)	39 (43.8)	8 (8.9)	4 (4.4)	0
TI	255 (73.9)	197 (77.2)	50 \pm 38	30 \pm 19	128 (50.1)	60 (23.5)	23 (38.3)	101 (39.6)	87 (34.1)	67 (26.2)	18 (7.0)	2 (0.7)	1 (0.3)

TM: Thalassemia Major, **TI:** Thalassemia Intermedia, **BT:** Blood Transfusion, **Xmn1:** -158 G \rightarrow T (Xmn1 is a restriction polymorphic site with known association with the severity of the disease)

Table 2: β -globin mutations in the β -thal patients

Mutation	HGVS nomenclature	Total allele (n)	Allele frequency (%)
IVS II-1 G\rightarrowA	HBB: c.315+1 G>A	353	40.33 %
IVS I-5 G\rightarrowC	HBB: c.92+5 G>C	83	9.56 %
CD30 G\rightarrowC	HBB: c.93 G>C	63	7.22 %
CD8-9 + G	HBB: c.27_28 ins G	61	7.00 %
IVS I-110 G\rightarrowA	HBB: c.93-21 G>A	43	4.89 %
CD 36-37 -T	HBB: c.112 del T	33	3.78 %
IVS I-6 T\rightarrowC	HBB: c.92+6 T>C	23	2.67 %
CD5 - CT	HBB: c.17_18 del CT	26	3.00 %
CD44 - C	HBB: c.135 delC	21	2.44 %
IVS I-1 G\rightarrowT	HBB: c.92+1G>T	17	1.89 %
-88 C\rightarrowT	HBB: c.-138 C>T	12	1.33 %
IVS I (-25 bp del)	HBB: c.93-21-96 del	10	1.11 %
CD 22 (-7 bp del)	----	22	2.56 %
Others (26 mutations)	-----	95	10.89 %
unknown	-----	4	0.46%
Total chromosome		866	100%

HGVS: Human Genome Variation Society, **IVS:** Intervening Sequence (Intron) (mutations in the introns of beta-globin are shown by IVS), **HBB:** Beta-globin gene, **CD:** Codon

Geographic distribution of population in Iran could be considered as the main reason of discordances between our study and other related researches.^{6, 17-18} No mutation was detected in 0.5% of β -thal alleles (n=4) neither by ARMS-PCR nor direct sequencing. However, two samples of them revealed anti 3.7 $\alpha\alpha\alpha$ through performing related PCR. Therefore, just a value of 0.25% of mutations could not be determined via the former methods. Association of Xmn1 and blood transfusion frequency has already been reported by Winichagoon et al.²⁸ Our research showed higher frequency of Xmn1 polymorphism (+/+ or +/-) in TI compared to TM patients (p<0.0001). Moreover, it is observed that non transfusion dependent thalassemia (NTDT) patients with β^0/β^0 genotype, inherited Xmn1 polymorphism (p<0.0001). Sivalingam et al.²⁹ have reported that patients carrying Xmn1 polymorphism required less frequent transfusion.

Dedousis et al.³⁰ have studied on the presence of clinical variability in patients who were homozygote or compound heterozygote for β^0 or β^+ thalassemia. They have found that rising fetal hemoglobin (HbF) level improved the clinical feature of the disease. Increasing of HbF has been attributed to the association of some β -globin mutations with the Xmn1 polymorphism. Pandey et al.²³ have reported

Table 3: Frequencies of different genotypes

	β^N/β^N	β^0/β^N	$\beta^0/\beta^{\text{unknown}}$	β^0/β^0	β^0/β^+	β^0/β^{++}	β^+/β^+	Hbvar/Hbvar
TM	0	0	1 (1.1%)	53 (59.5%)	11 (12.3%)	2 (2.2%)	12 (13.4%)	1 (1.1%) ($\delta\beta/\delta\beta$)
TI	1 (0.3%) (H-dis)	2 (0.7%) (anti-3.7)	1 (0.3%)	180 (71.3%)	39 (15.2%)	2 (0.7%)	26 (10.1%)	2 (0.7%) (HbS/HbS)

TM: Thalassemia major, **TI:** Thalassemia Intermedia, **HbVar:** Hemoglobin variant

that HbS- β thalassemia patients were clinically variable, ranging from a completely asymptomatic to a severe disorder. Others suggested that this heterogeneity could be caused by either different β -thal mutations or interactions between different genetic modulating factors like co-existence of α -thal and/or Xmn1 polymorphism.³¹

It has been reported that co-inheritance of α -thal could improve the clinical severity in β -thal patients.³² It is not possible to confirm that through this study. This discordance could be due to the lower frequency (10%) of α -globin gene deletions detected in the individuals. Different α -globin gene mutations have been shown to be prevalent in Iran accounting up to 30%,⁹ among them, α -globin gene deletions account for more than 60% of α -globin mutations.¹⁴⁻¹⁶ It was surprising to find lesser frequencies of α -deletions in our individuals.

Based on the results of this research, genotype determination is beneficial for early prognosis of β -thal and to choose the best possible treatment. Moreover, Xmn1 polymorphism has shown more ameliorating effect on the phenotype of our patients.

ACKNOWLEDGEMENT

Sincere thanks to technical staff of Kawsar Human Genetics Research Center for expert and amicable assistance. The authors express their gratitude to the referring physicians and the patients for their cooperation. This study was partly supported by the Iranian National Science Foundation (INSF), grant number: 86022/14 which we are thankful.

CONFLICT OF INTEREST

Authors declare that there is no conflict of interest.

REFERENCES

- Rahimi Z. Genetic epidemiology, hematological and clinical features of hemoglobinopathies in iran. *Biomed Res Int.* 2013; 2013:803487.
- Chaisue C, Kitcharoen S, Wilairat P, et al. Alpha/beta-Globin mRNA ratio determination by multiplex quantitative real-time reverse transcription-polymerase chain reaction as an indicator of globin gene function. *Clin Biochem.* 2007 Dec; 40(18):1373-7.
- Weatherall D.J, Clegg J.B. *The Thalassemia Syndromes.* 4th ed. Oxford: Malden/MA; Blackwell science, 2001:486, 192-247.
- Samavat A, Modell B. Iranian national thalassaemia screening programme. *BMJ.* 2004 Nov 13; 329(7475): 1134-7.
- Fallah MS, Samavat A, Zeinali S. Iranian national program for the prevention of thalassemia and prenatal diagnosis: mandatory premarital screening and legal medical abortion. *Prenat Diagn.* 2009 Dec; 29(13):1285-6.
- Abolghasemi H, Amid A, Zeinali S, et al. Thalassemia in Iran: epidemiology, prevention, and management. *J Pediatr Hematol Oncol.* 2007 Apr; 29(4):233-8.
- Hadipour Dehshal M, Tabrizi Namini M, Ahmadvand A, et al. Evaluation of the national prevention program in iran, 2007-2009: the accomplishments and challenges with reflections on the path ahead. *Hemoglobin.* 2014; 38(3):179-87.
- Galehdari H, Salehi B, Pedram M, et al. High prevalence of rare mutations in the Beta globin gene in an ethnic group in iran. *Iran Red Crescent Med J.* 2011 May; 13(5):356-8.
- Mahdavi MR, Kowsarian M, Karami H, et al. Prevalence of hemoglobin alpha-chain gene deletion in neonates in North of Iran. *Eur Rev Med Pharmacol Sci.* 2010 Oct; 14(10):871-5.
- Denic S, Agarwal MM, Al Dabbagh B, et al. Hemoglobin A2 Lowered by Iron Deficiency and alpha-Thalassemia: Should Screening Recommendation for beta-Thalassemia Change? *ISRN Hematol.* 2013; 2013:858294.

11. Alkindi SS, Alzadjali S, Daar S, et al. A stepwise alpha-thalassemia screening strategy in high-prevalence areas. *Eur J Haematol.* 2013 Aug; 91(2):164-9.
12. Saleh-Gohari N, Khosravi-Mashizi A. Spectrum of alpha-globin gene mutations in the Kerman Province of Iran. *Hemoglobin.* 2010; 34(5):451-60.
13. Rahim F. Correlation of beta-thalassemia mutations with alpha-thalassemia: an experience of the southwestern region of Iran. *Hematology.* 2010 Dec; 15(6):430-3.
14. Zandian K, Nateghi J, Keikhaie B, et al. Alpha-thalassemia mutations in Khuzestan Province, Southwest Iran. *Hemoglobin.* 2008;32(6):546-52.
15. Tamaddoni A, Hadavi V, Nejad NH, et al. Alpha-Thalassemia mutation analyses in Mazandaran province, North Iran. *Hemoglobin.* 2009; 33(2):115-23.
16. Hadavi V, Jafroodi M, Hafezi-Nejad N, et al. Alpha-thalassemia mutations in Gilan Province, North Iran. *Hemoglobin.* 2009; 33(3):235-41.
17. Mehrabi M, Alibakhshi R, Fathollahi S, et al. The spectrum of beta-thalassemia mutations in Kermanshah Province in West Iran and its association with hematological parameters. *Hemoglobin.* 2013; 37(6): 544-52.
18. Rahiminejad MS, Zeinali S, Afrasiabi A, et al. Beta-Thalassemia mutations found during 1 year of prenatal diagnoses in Fars Province, Iran. *Hemoglobin.* 2011; 35(4):331-7.
19. Yatim NF, Rahim MA, Menon K, et al. Molecular characterization of alpha- and beta-thalassaemia among Malay patients. *Int J Mol Sci.* 2014; 15(5):8835-45.
20. El-Shanshory M, Hagag A, Shebl S, et al. Spectrum of Beta Globin Gene Mutations in Egyptian Children with beta-Thalassemia. *Mediterr J Hematol Infect Dis.* 2014; 6(1): e2014071.
21. Nagar R, Sinha S, Raman R. Haemoglobinopathies in eastern Indian states: a demographic evaluation. *J Community Genet.* 2014 Jul 25.
22. Dadheech S, Jain S, Madhulatha D, et al. Association of Xmn1 -158 gammaG variant with severity and HbF levels in beta-thalassemia major and sickle cell anaemia. *Mol Biol Rep.* 2014 May; 41(5):3331-7.
23. Pandey S, Mishra RM, Saxena R. Modulating Effect of the -158 gamma (C-->T) Xmn1 Polymorphism in Indian Sickle Cell Patients. *Mediterr J Hematol Infect Dis.* 2012; 4(1):e2012001.
24. Miller SA, Dykes DD, Polesky HF. A simple salting out procedure for extracting DNA from human nucleated cells. *Nucleic Acids Res.* 1988 Feb 11; 16(3):1215.
25. Chong SS, Boehm CD, Higgs DR, et al. Single-tube multiplex-PCR screen for common deletional determinants of alpha-thalassemia. *Blood.* 2000 Jan 1; 95(1): 360-2.
26. Dode C, Krishnamoorthy R, Lamb J, et al. Rapid analysis of -alpha 3.7 thalassaemia and alpha alpha alpha anti 3.7 triplication by enzymatic amplification analysis. *Br J Haematol.* 1993 Jan; 83(1):105-11.
27. Sutton M, Bouhassira EE, Nagel RL. Polymerase chain reaction amplification applied to the determination of beta-like globin gene cluster haplotypes. *Am J Hematol.* 1989 Sep; 32(1):66-9.
28. Winichagoon P, Fucharoen S, Chen P, et al. Genetic factors affecting clinical severity in beta-thalassemia syndromes. *J Pediatr Hematol Oncol.* 2000 Nov-Dec; 22(6):573-80.
29. Sivalingam M, Looi ML, Zakaria SZ, et al. Molecular study and genotype/phenotype correlation of beta Thalassemia in Malaysia. *Int J Lab Hematol.* 2012 Aug; 34(4):377-82.
30. Dedoussis GV, Mandilara GD, Boussiu M, et al. HbF production in beta thalassaemia heterozygotes for the IVS-II-1 G-->A beta(0)-globin mutation. Implication of the haplotype and the (G)gamma-158 C-->T mutation on the HbF level. *Am J Hematol.* 2000 Jul; 64(3):151-5.
31. Serjeant G. *Sickle Cell Disease 3rd edition* ed: Oxford University press; 2001.
32. Nuntakarn L, Fucharoen S, Fucharoen G, et al. Molecular, hematological and clinical aspects of thalassemia major and thalassemia intermedia associated with Hb E-beta-thalassemia in Northeast Thailand. *Blood Cells Mol Dis.* 2009 Jan-Feb; 42(1):32-5.